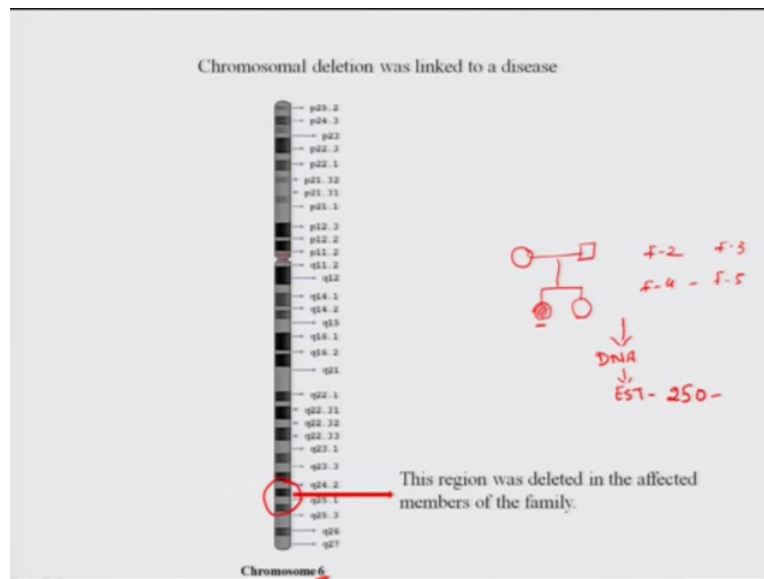


Functional Genomics
Professor S Ganesh
Department of Biological Sciences & Bioengineering
Indian Institute of Technology Kanpur
Lecture No 13
Tutorial Part 1

Hello every one and welcome to the tutorial session of functional genomics myself Anupama and as already introduced I am one of the teaching assistance in this course that is functional genomics today we will be learning about whatever information can be arrived at or fetched using few hundred base pairs of DNA sequence.

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So now let us look into that so this is a part of chromosome which was deleted in a disease and linked to that disease. This was identified by a team of scientists so few of you must be wondering that how did they arrived at the region of the chromosomes. So that has been covered already in one of the NPTEL course that is Human Molecular Genetics and also we are rerunning that course but since you are here very briefly I will like to tell you that how do we arrive at such sequences, so what is done that there are a families which come for counseling to the geneticists and they have members who are affected suppose this female was affected and she has a another sister and their parents were unaffected.

Similarly there was a another family, family 2, family 3 with family 4 so when you once you have a certain amount of family for which you can screen, screen for a cause of the disease. So what is done that DNA is extracted from all the family members which are present and then they are EST markers. Now what are EST markers these are small expressed sequence tag, which are unique for particular region of the chromosome.

These are unique for the regions so what do the scientists do they screen for these EST sequence, so suppose approximately around 250 EST sequence can cover almost all the autosomes of the chromosomes and from there on they link, they try to see that what EST region or what EST is getting segregated with the disease by that I mean that which one are always present in the individuals which are affected.

For example in this female and then the other person of the other family which is affected if that EST marker is present or not. So from that they narrowed down to the region of the chromosome and in this disease the case was the chromosom6 and this region, so from here what using this EST and a small probe they designed and then they screened then they screened a cDNA libraries for that.

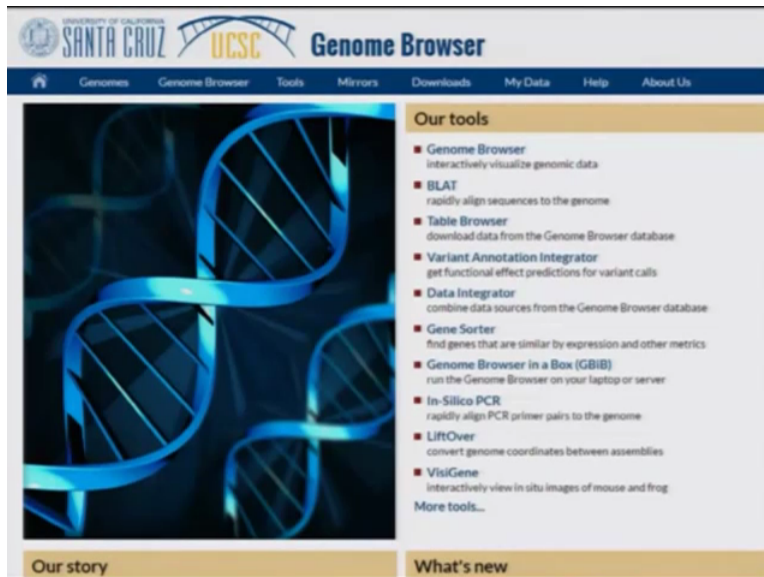
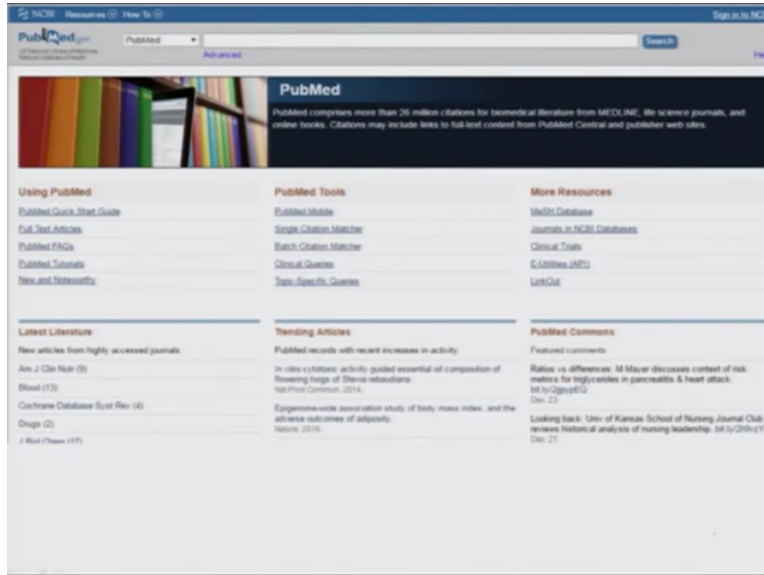
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They try to align them as they screen for many libraries sequence them as sequencing has already being taught and there are various ways by which you can sequence and then you arrive at a

sequence, in this session we will start with the sequence which has been arrived and they have few hundred base pair of the sequence, this is the 5 prime end and this is the 3 prime end of the sequence, so using the current search tools whatever information we can take is we can arrive at or which can be fetched using this few hundred base pair of the sequence that is what we will be learning today.

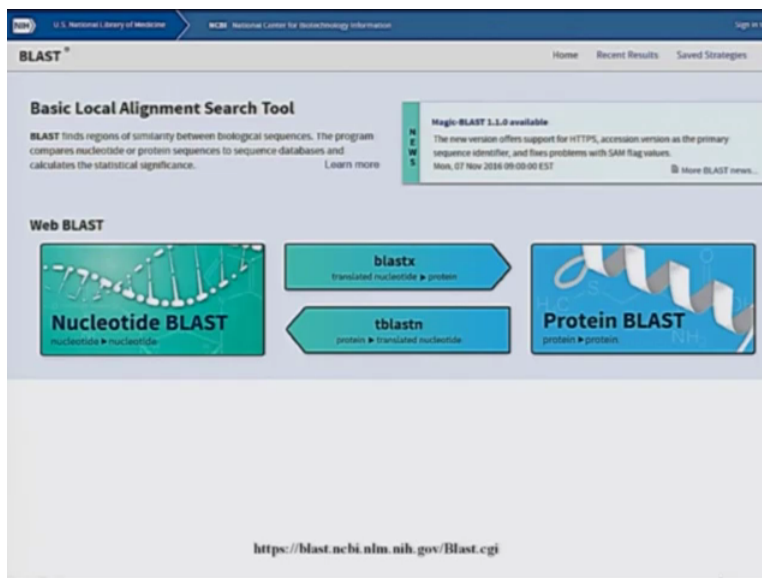
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So using there are few widely used search engines one of those is NCBI another one is UCSC genome browser another one is Ensembl, so today we will be learning about more about NCBI and then we will look online on one of the another search engine that is Ensembl. So since we had a few hundred base pair of sequence and we did not knew from which gene it is or whether this is a coding region or not coding region or this is just a intron of a gene or this a exon of a gene all those information we do not have.

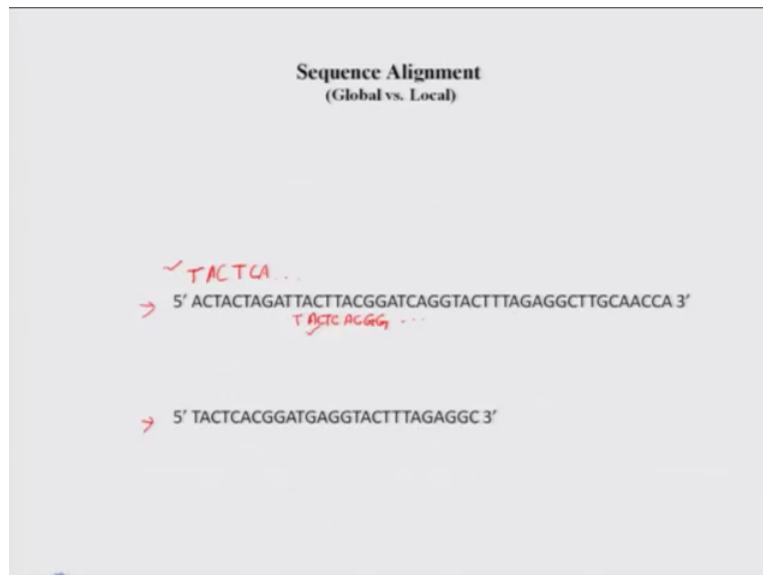
(Refer Slide Time: 5:05)



So one such way is to use blast and here is the page of blast that is that is Basic Local Alignment Search Tool. So what does blast does it finds the region of similarity between biological sequences, this programme it compares the nucleotides or protein sequences to sequence data base and calculate the statistical significance, so they are various blast programmes like one is nuclear type blast, the other one is protein blast. Now you have a stretch of a sequence that can be converted into protein and then again it can be blast to the protein data base that is called blastx.

Similarly the protein can be translated to nucleotide and then it can be blast to the other nucleotide data base and that is called tblastn, so as we have seen that we had a DNA sequence so we will go for nucleotide blast and try to look that what region it is aligning to but before we go into what region it is aligning to very briefly we will look into what does sequence alignment means and what are the various strategies used to align the sequence.

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So there are two key terms which you will encounter more often that is global versus local alignment so very briefly what is global and what is local alignment. Let us look into these two sequences now one you have to align these two sequences one is the one this sequence and the other one is this. Now there are two ways by two ways by which you can align this sequence one is that you just go one by one and try and align for example here if I align this one with this so this would be TACTCA and so on.

And you will try and calculate this core which is based on the various algorithms that what is this core of this alignment, now another way is to that you leave you do not go one by one but you will try and look for small stretch which are aligning the best for example here it would be if you align here with this TACTCACGG and so on you will see that the local stretches are aligning more, so this is called global alignment the first method and the other method is called the local alignment.

Similarly if you have to align these two sequences which are almost of same lengths then it is best to align them globally. So when there is a small stretch of sequence its best to align them local and if there are equal length of equal length of DNA sequences then that can be aligned globally. So on these alignments using different algorithms scores are calculated, few of those are like from where you are starting, what is the gap between the first not aligned and the one where the alignment starts there is a gap penalty then in between there are gap penalty where there are mismatches or you skip few bases and then you align to another stretch.

For example suppose you this is one of this sequence this is quite frank 3 prime and then you have another sequence so it aligns here but then there are some mismatches so you skip that and then again you align. So there would be a certain penalty for this and then that penalty would increase at these regions so on basis of the score is calculated and that is also represented in a blast data.

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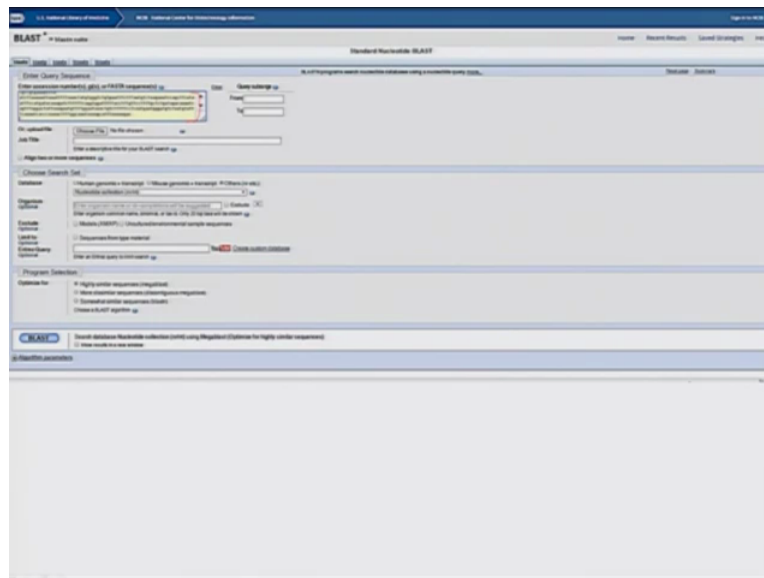
The image shows the NCBI BLAST Standard Nucleotide BLAST web interface. At the top, it displays the NCBI logo and navigation links like 'Home' and 'Recent Results'. The main heading is 'BLAST -> blastn suite' and 'Standard Nucleotide BLAST'. Below this, there are tabs for 'blastn', 'blastp', 'blastx', 'tblastn', and 'tblastx'. The 'Enter Query Sequence' section includes a text input field for accession numbers, a 'Clear' button, and a 'Query subrange' section with 'From' and 'To' fields. There is also an option to 'Or, upload file' with a 'Choose File' button. The 'Job Title' section has a text input field. Below that, there is a checkbox for 'Align two or more sequences'. The 'Choose Search Set' section includes a 'Database' dropdown menu (currently set to 'Nucleotide collection (nr/nt)'), an 'Organism' text input field, and several checkboxes for 'Exclude' options: 'Models (DMXP)', 'Uncultured/environmental sample sequences', and 'Sequences from type material'. There is also an 'Enter Query' field for limiting the search.

So now let us look into how do we go about it, so what happens like this is the blast home page where you have different ways as I have already described blastn, blastp for protein blastx where nucleotide is converted to the protein and then blast to the whole protein data base.

Similarly the nucleotide is translated and then it is blasted to or aligned to to the sequences in the protein data base and so on, so here you can enter your sequence which has that sequence can be entered in a different formats which has accretion numbers which are given by NCBI and that is at the unique suppose you have a gene for which you want to blast, you just need to put that accession number and the tool would take it or you can paste faster sequence.

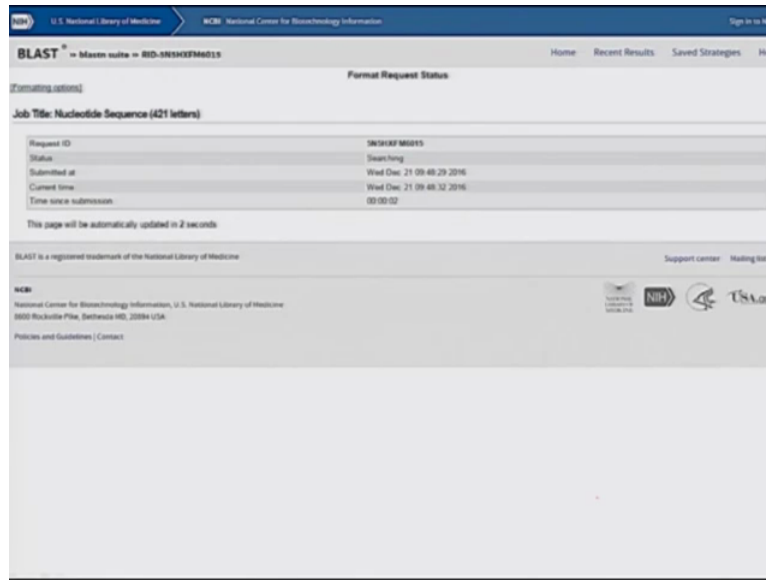
Now faster sequence is a format of writing the nucleotide or the DNA sequences then on the basis of your of your requirement you can make the... You can use a particular data base suppose you just want to use human the genomics or the transcript or mouse genomics transcript or you can choose from other organism or exclude them add them whatever is your criteria you can do that and then you can just click to blast.

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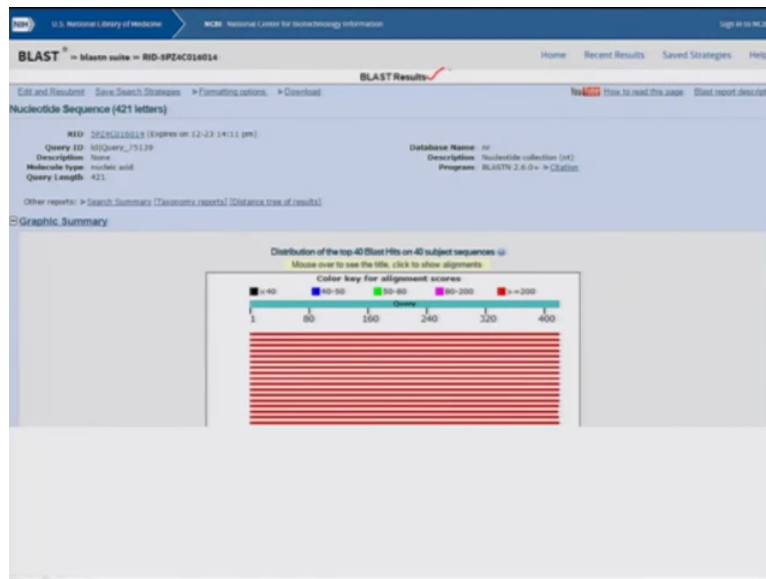
So in our case this was the query sequence which I have pasted here and then you click on blast just this is the default setting. I have not changed anything and you click on blast.

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So what do you get the results sometimes the software takes time and doing that and this is how the page it comes. This is the job title and this is your query sequence and this is the time you submitted and how much time duration has passed on.

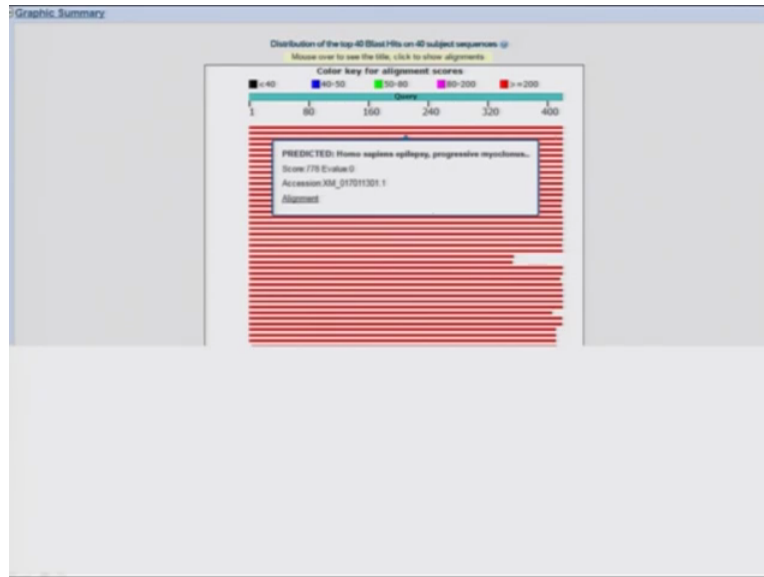
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Then you have a result place this is the blast result page so here are the this is the query the description you had put in nucleic acid which was 421 base pair long and this is the data base through which it has done the search and that is non-redundant data base. Non redundant data

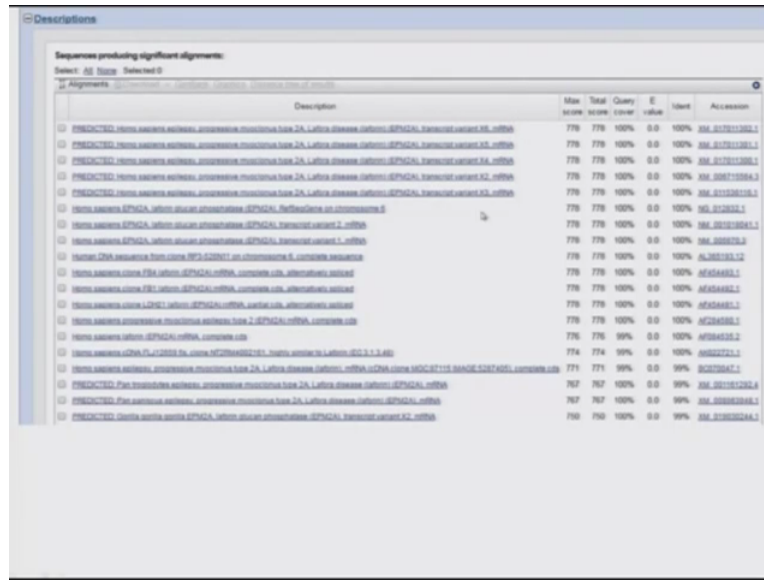
base then it has the description as the nucleotide collections then this is blastn and then you get a distribution of the top 40 blast on the 40 subject sequence, so these are the 40 sequence subjects, subjects sequences against which these are the best scored ones and the score has been here shown in the format of colour. So red shows the highest one and the black one black shows the lowest one and this is your query sequence which was 421 base pair long.

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Now in this you can always get your mouse there and you can find that what that sequence is what is the alignment, what is the accession number and what and what is the E values score which I will be explaining in a short duration.

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The screenshot shows a BLAST search results page titled "Descriptions". It displays a list of sequences that have aligned with the query. The table has the following columns: Description, Max score, Total score, Query cover, E value, Ident, and Accession. The first few rows show sequences with a Max score of 778, Total score of 778, Query cover of 100%, and E value of 0.0. The Accession numbers for these sequences are: XM_017211282.1, XM_017211281.1, XM_017211280.1, XM_008712884.3, and XM_012281282.1. The table continues with more sequences, including those from the Homo sapiens EPNCA gene and other species like Rattus norvegicus and Mus musculus.

Description	Max score	Total score	Query cover	E value	Ident	Accession
PR000332 Homo sapiens EPNCA, transcript variant X6, mRNA	778	778	100%	0.0	100%	XM_017211282.1
PR000332 Homo sapiens EPNCA, transcript variant X5, mRNA	778	778	100%	0.0	100%	XM_017211281.1
PR000332 Homo sapiens EPNCA, transcript variant X4, mRNA	778	778	100%	0.0	100%	XM_017211280.1
PR000332 Homo sapiens EPNCA, transcript variant X2, mRNA	778	778	100%	0.0	100%	XM_008712884.3
PR000332 Homo sapiens EPNCA, transcript variant X3, mRNA	778	778	100%	0.0	100%	XM_012281282.1
Homo sapiens EPNCA, RefSeqGene on chromosome 6	778	778	100%	0.0	100%	NC_017202.1
Homo sapiens EPNCA, transcript variant 7, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 1, mRNA	778	778	100%	0.0	100%	XM_00022281.2
Rattus norvegicus EPNCA, transcript variant 1, mRNA	778	778	100%	0.0	100%	XM_00022281.2
Homo sapiens EPNCA, transcript variant 2, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 3, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 4, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 5, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 6, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 8, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 9, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 10, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 11, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 12, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 13, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 14, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 15, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 16, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 17, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 18, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 19, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 20, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 21, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 22, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 23, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 24, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 25, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 26, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 27, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 28, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 29, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 30, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 31, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 32, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 33, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 34, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 35, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 36, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 37, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 38, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 39, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 40, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 41, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 42, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 43, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 44, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 45, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 46, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 47, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 48, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 49, mRNA	778	778	100%	0.0	100%	XM_00222281.1
Homo sapiens EPNCA, transcript variant 50, mRNA	778	778	100%	0.0	100%	XM_00222281.1

So this is our description page where we have seen that our query sequence has aligned to these many sequences and there were many more however through the print screen I have got only this one this much. But when you will be doing it online you can scroll down and see that there were many more sequences from various species to which your queries sequence has aligned so what could be the maximum score the total score and how much the query has been covered is given here.

Now what is E value so E value defines the number of hits which you can expect by chance, so that here it is zero which means that all the base pairs of your query were aligning hundred percent and it was not a chance event and here you have accession numbers by which you can access this particular transcript which is here it is like EPM2A which is associated with I suppose lafora disease and the protein is laforan and it is a transcript variant X6 mRNA and you can access these by this accession ID.

(Refer Slide Time: 13:48)

The screenshot displays a BLAST search interface. At the top, it identifies the predicted sequence as "Homo sapiens epilepsy, progressive myoclonus type 2A, Lafora disease (laforin) (EPM2A), transcript variant X6, mRNA" with accession number XM_017011302.1. Below this, a table provides summary statistics for the alignment: Score: 778, Expect: 0.0, Identities: 431/431 (100%), Gaps: 0/42 (0%), and Pos/Phy: 431/431. The main section shows the alignment of the query sequence (Query 1) against the subject sequence (Sbjct 2393). The alignment starts at position 2393 and continues until position 2813, with the query sequence being 421 nucleotides long. The alignment is shown as a series of vertical bars representing matches between the query and subject sequences.

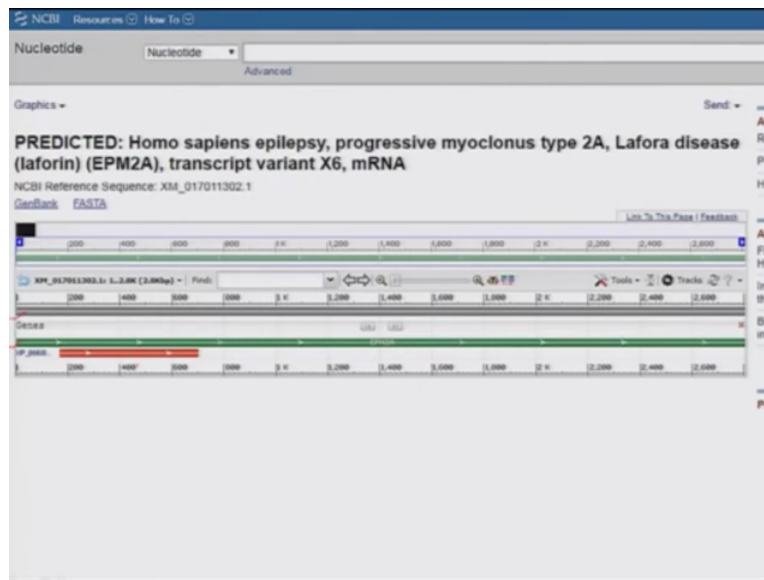
So once you have that this accession ID then you can also look into how your query sequence was aligning to the subject sequence or to this sequence which the search engine has searched for. So here you see that your this is the query and this is the subject sequence. Your query first nucleotide is aligning with with the 2393 or 2393 base pair of the subject sequence and from there on it is aligning to till 2813 base pair.

(Refer Slide Time: 14:28)

The screenshot shows the NCBI GenBank record for the predicted sequence XM_017011302.1. The record includes the following information:
- **LOCUS:** XM_017011302 2813 bp mRNA linear PRI 06-JUN-2016
- **DEFINITION:** PREDICTED: Homo sapiens epilepsy, progressive myoclonus type 2A, Lafora disease (laforin) (EPM2A), transcript variant X6, mRNA.
- **ACCESSION:** XM_017011302
- **VERSION:** XM_017011302.1
- **DBLINK:** BioProject: PRJNA448
- **KEYWORDS:** RefSeq
- **SOURCE:** Homo sapiens (human)
- **ORGANISM:** Homo sapiens
- **COMMENT:** MODEL REFSEQ: This record is predicted by automated computational analysis. This record is derived from a genomic sequence (NC_000006.12) annotated using gene prediction method: Gnomon, supported by mRNA and EST evidence. Also see: Documentation of NCBI's Annotation Process
- **##Genome-Annotation-Data-START##**
- **Annotation Provider:** NCBI
- **Annotation Status:** Full annotation
- **Annotation Version:** Homo_sapiens_Annotation_Release_108
- **Annotation Pipeline:** NBT eukaryote genome annotation
The right-hand side of the page contains a sidebar with various tools and links, including "Analyze this", "Run BLAST", "Pick Primer", "Highlight SNPs", "Find in this genome", "Articles at PubMed", "FH Regular Expression", "Interdependent stable", "Biophysical", "carbohydrate", "Pathways", "Glycogen", and "Glucose".

Now from here you can access the sequence what do you find that this the first hit which the search engine has given, it is a protein EPM2A or laforan EPM2A or Laforan and it is associated with a progressive myoclonus type of epilepsy. The below you have all the information about this entry in whatever organisms it is found it is found in the (())(14:49) cod data then there are commons about it. What genome annotation data it has been provided for example from NCBI the annotation status and then as you will scroll down you will get lots of information about that.

(Refer Slide Time: 15:23)



If you click on the graphics of this you can see the alignment in the form of a graph here this is your query sequence and this is EPM2A and how they are aligning. You can always explore going to these arrows or you can use the tools and tracks and whatever you can explore all this online.

(Refer Slide Time: 15:41)

The screenshot shows the NCBI Nucleotide database interface. The main content area displays the predicted sequence for Homo sapiens epilepsy, progressive myoclonus type 2A, Lafora disease (laforin) (EPM2A), transcript variant X6, mRNA. The sequence is shown in FASTA format, with a red vertical line highlighting a specific region. The right sidebar contains several sections: 'Change region shown', 'Customize view', 'Analyze this sequence' (with options for Run BLAST, Pick Primers, Highlight Sequence Features, and Find in this Sequence), 'Articles about the EPM2A gene' (with links to 'F0H Regulates Cellular Metabolism through Hydroxylation of the Dodecyls (PLoS Biol. 2010)', 'Interdependence of laforin and main proteins for their stability and function (codr11 Biosci. 2010)', and 'Biophysical characterization of laforin catalytic-site interaction. (Biochem J. 2010)'), and 'Pathways for the EPM2A gene' (with links to 'Glycogen synthesis' and 'Glucose metabolism').

Similarly then there you can go for the FASTA sequence and you will find the sequence of the gene to which your query has aligned at the most, from here as you can see on the right hand side there are various ways to analyse these sequences for example you can run a blast in which this now this sequence of the subject which you have arrived at that is EPM2A gene or lafora can be used to used to run a blast against all the nucleotide sequence.

Similarly you can use primers to amplify this particular gene you can highlight the sequence speeches what are the characteristics sequence speeches and you can also find something in this sequence suppose you are looking for a particular stretch of a sequence you can use this tool to identify, similarly there are articles about EPM2A gene as you are trying to find out the gene for the linked disease so you have come so far so you can always look into what are the recent articles related to with gene, what are the pathways in which this genes is involved like for example glycogen , synthesis or glucose metabolism.

You can always from here link to the phenotype of the disease is this are theses pathways defective in those patience in which this are theses chromosomal region is deleted. Do you see something abrupt with glycogen metabolism in those patients? So with this you can link the information in the tool.

(Refer Slide Time: 17:20)

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BLAST® » blastn suite

Standard Nucleotide BLAST

BLASTN programs search nucleotide databases using a nucleotide query. 2025...

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s)

Or, upload file No file chosen

Job Title

Align two or more sequences

Choose Search Set

Database Human genomic + transcript Mouse genomic + transcript Others (w/ etc.)

Nucleotide collection (nr/nt)

Organism

Exclude Models (XM/XP) Uncultured/environmental sample sequences

Limit to Sequences from tree material

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BLAST® » blastn suite » RID-3RAHF15Z015

Format Request Status

[Formatting options]

Job Title: ref|XM_017011302.1| (2813 letters)

Request ID	SRAHF15Z015
Status	Searching
Submitted at	Thu Dec 22 05:25:51 2016
Current time	Thu Dec 22 05:25:59 2016
Time since submission	00:00:07

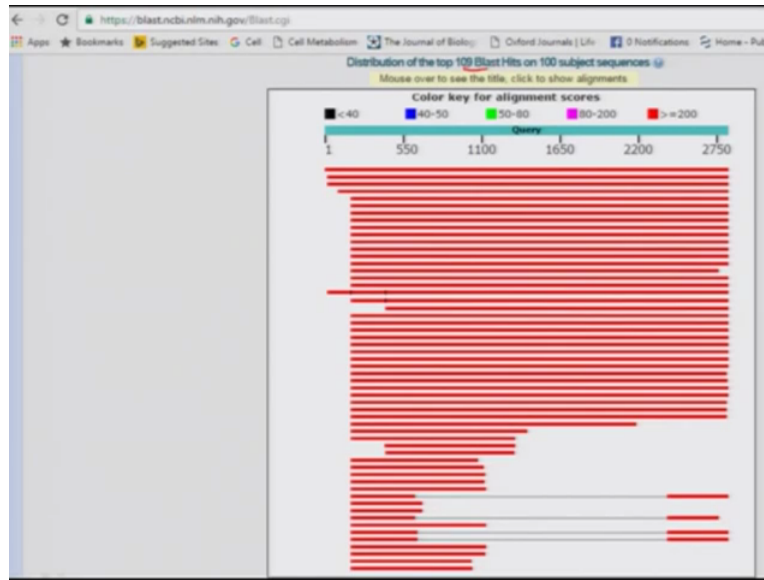
This page will be automatically updated in 2 seconds

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NCBI
National Center for Biotechnology Information, U.S. National Library of Medicine

When you run blast using that query now again this is just taken the accession number. As I have already described that anything you can use as a query sequence in the NCBI blast and then you can again using the default as I have done here you can run blast again the search engine takes time and thus the page appears on your screen and then this is the way the result is again.

(Refer Slide Time: 17:54)



So this is the similar result as I have described so here these are top 109 blast hits on the hundred subject sequences and this is how it is aligning mostly are red almost all are red and these are the stretch of sequences now these are the stretch of sequences which were not aligning and the last part was aligning.

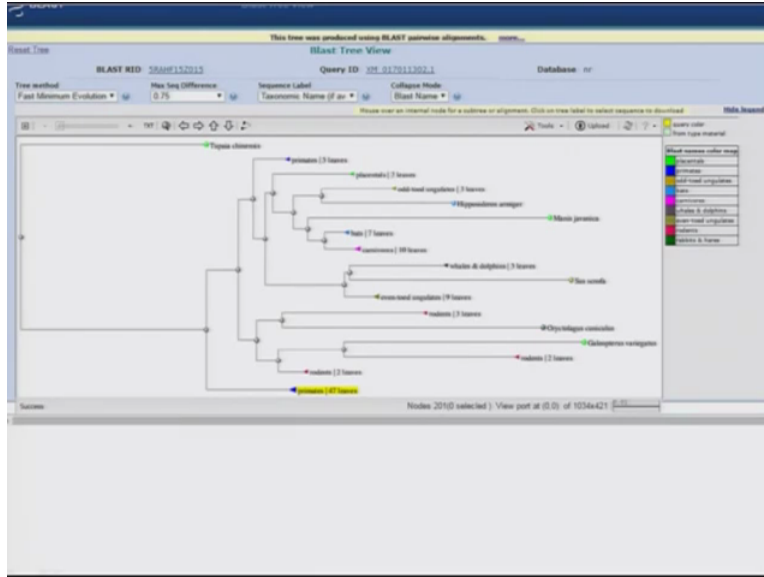
(Refer Slide Time: 18:14)

Description	Max score	Total score	Query cover	E value	Identities	Acc
PF00730 Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A), transcript variant 10, mRNA	5196	5196	100%	0.0	100%	XM_01
PF00730 Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A), transcript variant 11, mRNA	5160	5160	99%	0.0	100%	XM_01
PF00730 Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A), transcript variant 14, mRNA	5160	5160	99%	0.0	100%	XM_01
Homo sapiens cDNA FL122539.1c clone NT264822.111, highly similar to Leifera (EPM2A)	5011	5011	96%	0.0	99%	AF5222
PF00730 Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A), transcript variant 13, mRNA	4861	4861	93%	0.0	100%	XM_01
Homo sapiens EPM2A, isoform alpha, alternative splicing (EPM2A), transcript variant 1, mRNA	4861	4861	93%	0.0	100%	XM_01
Homo sapiens clone F84, isoform (EPM2A) mRNA, complete cds, alternative splicing	4841	4841	93%	0.0	99%	AF5544
Homo sapiens clone F81, isoform (EPM2A) mRNA, complete cds, alternative splicing	4841	4841	93%	0.0	99%	AF5544
Homo sapiens ectopic progressive mucosus type 2 (EPM2A) mRNA, complete cds	4841	4841	93%	0.0	99%	AF5544
Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) mRNA cDNA (EPM2A) clone M30387.11 IMAGE 5287451, complete cds	4815	4815	93%	0.0	99%	BC1222
Homo sapiens clone L2421, isoform (EPM2A) mRNA, partial cds, alternative splicing	4809	4809	93%	0.0	99%	AF5544
Homo sapiens isoform (EPM2A) mRNA, complete cds	4798	4798	93%	0.0	99%	AF5544
PF00730 Pan paniscus ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A) mRNA	4721	4721	93%	0.0	99%	XM_01
PF00730 Pan troglodytes ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A) mRNA	4704	4704	93%	0.0	99%	XM_01
Homo sapiens mRNA for EPM2A, isoform 1, partial	4704	4704	91%	0.0	99%	AF1220
PF00730 Gorilla gorilla gorilla EPM2A, isoform alpha, alternative splicing (EPM2A), transcript variant 12, mRNA	4645	4645	93%	0.0	99%	XM_01
PF00730 Gorilla gorilla gorilla EPM2A, isoform alpha, alternative splicing (EPM2A), transcript variant 13, mRNA	4645	4645	93%	0.0	99%	XM_01
Homo sapiens EPM2A, isoform alpha, alternative splicing (EPM2A), RefSeqGene on chromosome 8	4423	5173	99%	0.0	99%	XM_01
Human cDNA sequence from clone SP3-170917 on chromosome 8, complete sequence	4423	4873	93%	0.0	99%	AF1220
PF00730 Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A), transcript variant 12, mRNA	4420	4420	88%	0.0	100%	XM_01
AF007479 Homo sapiens ectopic progressive mucosus type 2A, Leifera disease (refseq) (EPM2A), transcript variant 11, mRNA	4368	4368	93%	0.0	99%	XM_01

So once you have the sequence or the all the sequence which are aligning to that gene that was EPM2A, EPM2A gene and these are the genes which are now aligning to that gene. You can

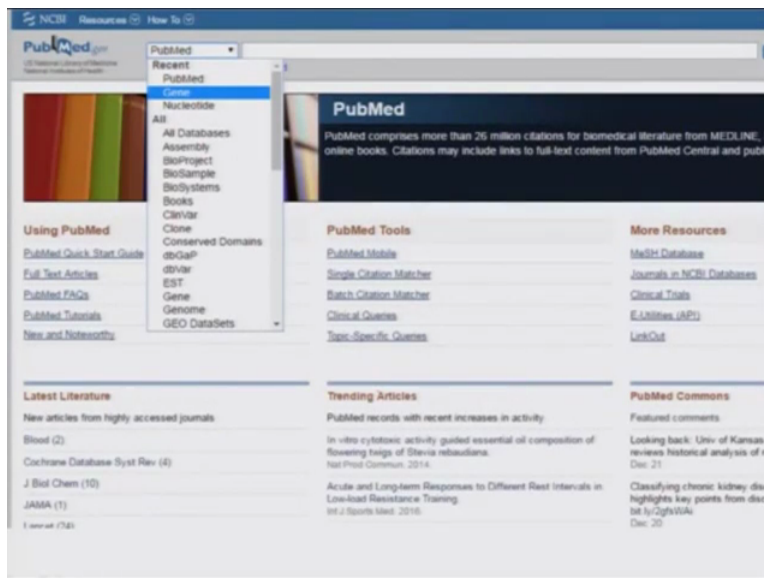
select all clicking on all and then you can go for distance tree results so from here you can get the phylogenetic tree of the gene of your interest.

(Refer Slide Time: 18:46)



So this tree shows how many how the gene has evolved and how close to one or the other organisms genes is how it is. So for example primates are more closer to rodents and so on, so once you have the sequence.

(Refer Slide Time: 19:03)



So you can also access to the protein of the sequence you can go to PubMed and you can go on gene.

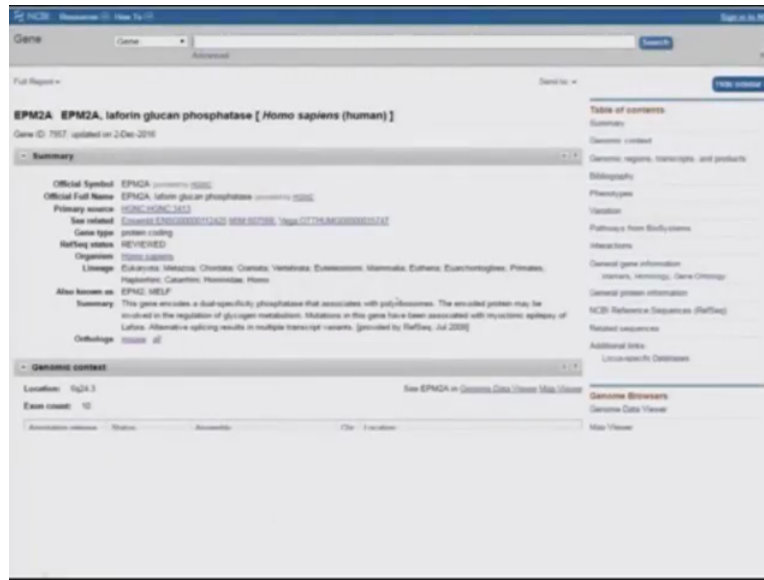
(Refer Slide Time: 19:16)

The screenshot shows the NCBI Gene database search results for the query 'EPM2A'. The search results are displayed in a table with columns for Name/Gene ID, Description, Location, Aliases, and MIM. The first result is EPM2A (ID: 7957) in Homo sapiens (human), located on Chromosome 6, NC_000006.12 (145500744-145736018, complement). Other results include Epm2a in Mus musculus (house mouse), Epm2a in Rattus norvegicus (Norway rat), epm2a in Danio rerio (zebrafish), and epm2a in Leposteus oculatus (spotted gar).

Name/Gene ID	Description	Location	Aliases	MIM
EPM2A ID: 7957	EPM2A, laforin glucan phosphatase [Homo sapiens (human)]	Chromosome 6, NC_000006.12 (145500744-145736018, complement)	EPM2, MELF	607566
Epm2a ID: 13883	epilepsy, progressive myoclonic; epilepsy, type 2 gene alpha [Mus musculus (house mouse)]	Chromosome 10, NC_000076.6 (11343445-11457477)	TG-B, Tg(TraK, TruK)TG-BFiv	
Epm2a ID: 114005	epilepsy, progressive myoclonus type 2A [Rattus norvegicus (Norway rat)]	Chromosome 1, NC_005100.4 (5448968-5571512)		
epm2a ID: 100535304	epilepsy, progressive myoclonus type 2A, Lafora disease (lafum) [Danio rerio (zebrafish)]	Chromosome 23, NC_007134.6 (1517485-1548489, complement)	si:ch1073-93a12.2	
epm2a ID: 102687876	EPM2A, laforin glucan phosphatase [Leposteus oculatus (spotted gar)]	Chromosome LG1, NC_023179.1 (25236514-25266287)		

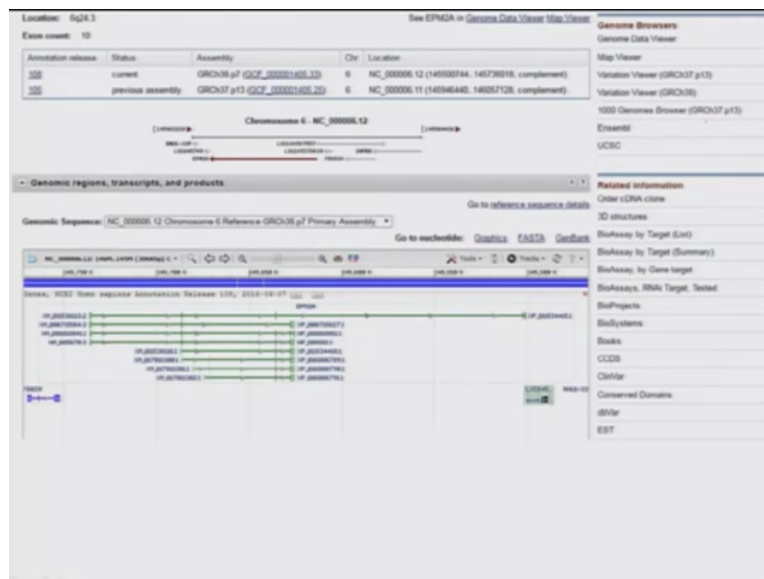
From here you will write the gene which you have identified that is EPM2A and then you can select whatever organisms you want for example you want the gene which is there sequenced in the humans or in the mouse or in the rat here we our query was about a sequence which we have identified from a part of a chromosome which was deleted in the family of of the affected people.

(Refer Slide Time: 19:45)



So we will go for humans and then you can have all the information about this gene in the humans from what is the lineage what is the summary what does this gene does what are the orthologs present.

(Refer Slide Time: 20:02)



Similarly from the on the right hand side you have all the other information then you can scroll down and look into what are the other cDNA clones, 3D structures and everything. What are the EST associated with this gene and so on, now here you see on the screen that this is showing

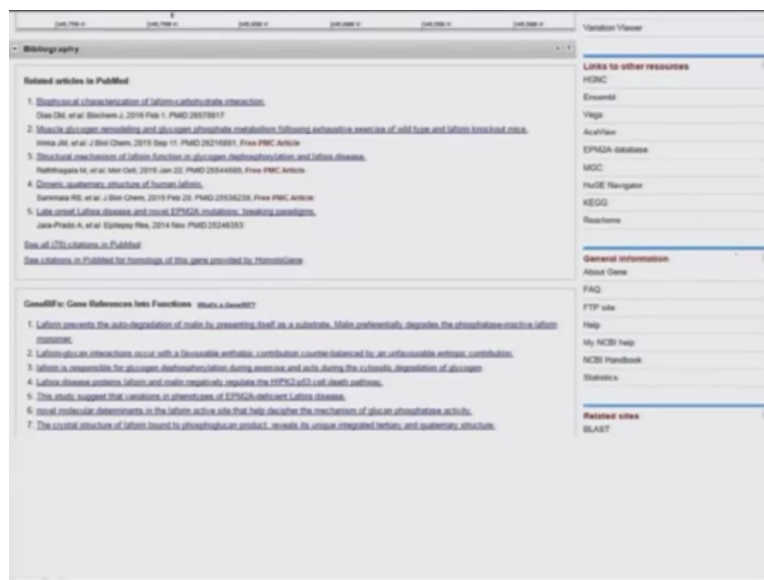
what are the transcripts present so there are I, 2 3, 4, 5, 6, 7 and 8, 8 transcripts for this gene that is EPM2A.

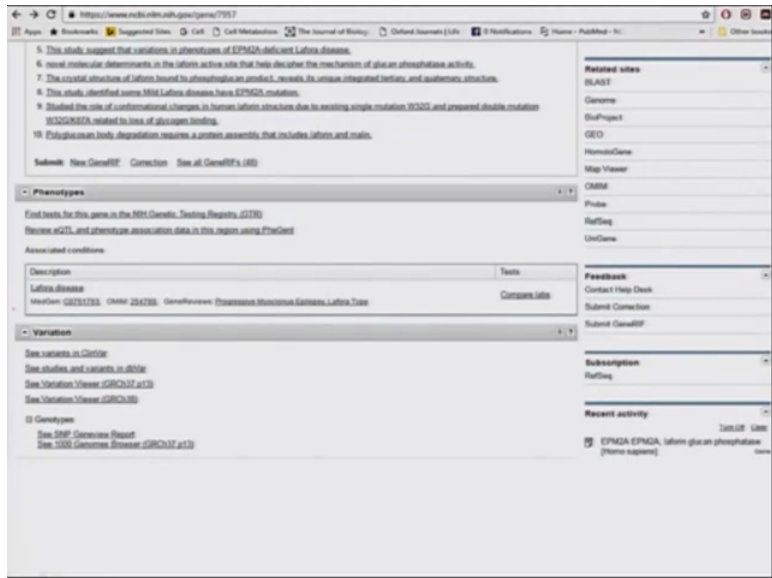
(Refer Slide Time: 20:38)



Then on again you can look for OMIM which is Online Mendelian Inheritance of Man that is this is another search tool by you can search for EPM2A and you can see that to what all disease it is associated with.

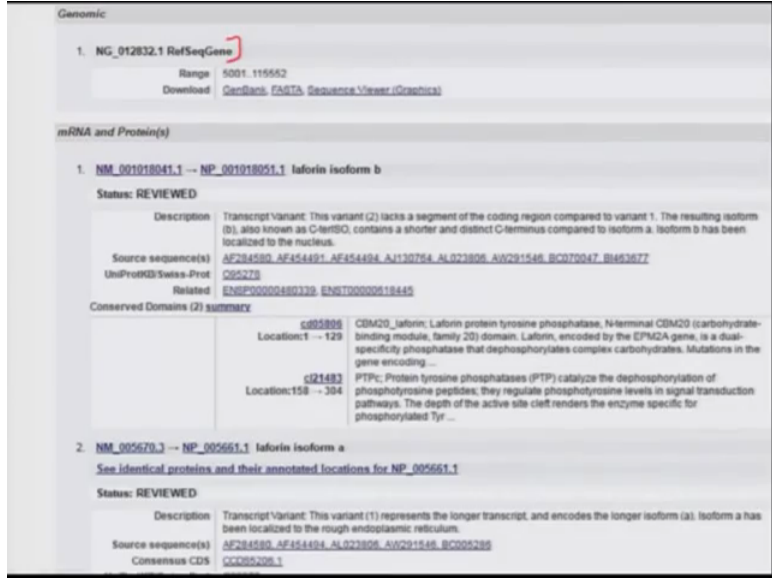
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Then again you have all the related articles and then gene reference and there is a huge amount of information already available on the internet which can help you arrive if all kind of information you are looking at but you need to know what you are what do you what exactly you want to search.

(Refer Slide Time: 21:21)



Similarly when you will scroll down you will see that there are the reference sequences of the gene you can go to the chromosomal region from which this gene was coded and there you can have all the information about introns, exons what are the sequences as the human chromosome

or the human genome has been sequenced aligned and deposited on internet for our ease. Similarly there is a protein sequence and mRNA sequence NM generally is for the mRNA sequence and NP is mainly for proteins but sometimes you will also see accession IDs with xp and all, these are for the proteins.

(Refer Slide Time: 22:17)

The screenshot displays the NCBI protein database entry for **laforin isoform b [Homo sapiens]**. The entry includes the following information:

- NCBI Reference Sequence:** NP_001018051.1
- Definition:** laforin isoform b [Homo sapiens]
- Accession:** NP_001018051
- Version:** NP_001018051.1
- Sequence:** RefSeq; accession [U000018051.1](#)
- Source:** Homo sapiens (human)
- Organism:** *Homo sapiens* (human); Chordata; Vertebrata; Euteleostomi; Mammalia; Eulalia; Euarchontoglires; Primates; Haplorhina; Catarrhini; Hominidae; Homo.
- References:** 1 (residues 1 to 337).
2 (residues 1 to 337).
- 3D Structure:** Structure of A Product Bound Phosphatase. PDB: 4R0X. Source: Homo sapiens. Method: X-Ray Diffraction. Resolution: 2.4 Å.
- Articles about the EPINCA gene:** Biophysical characterization of laforin-catalyzed interactions. [Biochem J. 2010].

So from here you can go to the protein sequence which this gene is coding for and then you have all the information about the protein data base that is you can have the conserve domain known you can highlight some sequence feature you can use the FASTA sequence and you can run a blast as in against all the protein data base and so on.

(Refer Slide Time: 22:38)

The screenshot shows the NCBI Protein database interface. The top navigation bar includes 'NCBI Resources' and 'How to'. Below the search bar, the protein name 'laforin isoform b [Homo sapiens]' is displayed. The FASTA sequence is shown in a monospaced font. On the right side, there are several interactive options: 'Change region shown', 'Analyze this sequence' (with a sub-option 'Run BLAST' checked), 'Identify Conserved Domains', 'Highlight Sequence Features', and 'Find in the Sequence'. Below these, there is a 'Protein 3D Structure' section with a small image of the protein structure and details: 'Structure Of A Product Bound Phosphatase', 'PDB: 4J6K', 'Source: Homo sapiens', 'Method: X-Ray Diffraction', and 'Resolution: 2.4 Å'. At the bottom, there are 'Articles about the EPN2A gene' with links to scientific papers.

So with this we have the FASTA sequence and we can run blast this protein is aligning to what all species of the protein sequences it is aligning to the protein sequences of what all species.

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This screenshot is identical to the one above, but with a blue rectangular box highlighting the text: 'View conserved domains detected in this protein sequence using CD-search'. This text is located in the middle of the FASTA sequence area, overlapping the sequence itself.

Conserved Domains

Conserved domains on [gi|66346728|ref|NP_001018051.1|]
laforin isoform b [Homo sapiens]

Protein Classification
CBM20, laforin and PTPc domain-containing protein (domain architecture ID 10146012)
CBM20, laforin and PTPc domain-containing protein

Graphical summary Zoom to residue level show extra options

Query seq.
 carbohydrate site 1 A
 carbohydrate site 2 B
 active site
 catalytic residues

Specific NID
 SuperFamilies **CBM20 superfamily** **PTPc superfamily**

List of domain hits

Name	Accession	Description	Interval	E-value
[H] CBM20, laforin	cd05006	Laforin protein tyrosine phosphatase, N-terminal CBM20 (carbohydrate-binding module, family 20)...	1-129	3.09e-05
[H] PTPc super family	cd21463	Protein tyrosine phosphatases (PTP) catalyze the dephosphorylation of phosphotyrosine peptides;...	158-204	2.55e-13

References:
 Hershler-Bauer A et al. (2015), "CDD: NCBI's conserved domain database.", *Nucleic Acids Res.* 43(D):D222-6.
 Hershler-Bauer A et al. (2011), "CDD: a Conserved Domain Database for the functional annotation of proteins.", *Nucleic Acids Res.* 39(D):D225-9.
 Hershler-Bauer A et al. (2009), "CDD: specific functional annotation with the Conserved Domain Database.", *Nucleic Acids Res.* 37(D):D209-16.
 Hershler-Bauer A, Bryant SH (2004), "CD-Search: protein domain annotations on the fly.", *Nucleic Acids Res.* 32(W):W327-331.

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Similarly you can identify conserve domains. So if you click here you will get to know what are the conserved domains or motives in this proteins sequence for example in this protein which was being coded by EPM2A it has CBM20, it is carbohydrate binding domain. So as the function of EPM2A was in glycogen metabolism and in glycogen synthesis path way so it will goes with the the functions which it does that it would bind to the starch or the glycogen, similarly there is another region that is PTPC super family. What is PTPC super family? It is a protein tyrosine phosphatase which catalyses the de-phosphorylation of phospho tyrosine peptide.

So this protein which was being encoded by EPM2A it has two domains one is CBM20 which has ability to bind two glycogen like molecules and similarly it has an another domain that is PTPC which is for the which is a phosphatase domain by that we mean that it can de-phosphorylate the phosphate group from the peptides.

(Refer Slide Time: 24:08)

The screenshot shows the NCBI Protein database interface for the entry 'laforin isoform b [Homo sapiens]'. The FASTA sequence is displayed, and a blue box highlights a feature bar within the sequence. A callout box explains: 'Open the Highlight Feature Bar and highlights feature annotations from the FEATURES table of the record. The Highlight Feature Bar can be used to navigate to and highlight other features and provides links to display the highlighted region separately. Links in the FEATURES table will also highlight the corresponding region of the sequence. Here...'. The right sidebar contains options to 'Change region shown', 'Analyze this sequence', 'Protein 3D Structure', and 'Articles about the EPMAA gene'.

Similarly you can highlight the sequence features of this peptide sequence.

(Refer Slide Time: 24:14)

The screenshot shows the NCBI Protein database interface for the entry 'laforin isoform b [Homo sapiens]'. The FASTA sequence is displayed, and the feature bar is highlighted in orange. The feature bar contains the following information: '1..337', '/gene="EPMAA"', '/gene_synonym="EPMA2; HELP"', '/coded_by="NP_001018051.1:158..1311"', '/note="isoform b is encoded by transcript variant 2"', '/db_xref="GeneID: 2322"', '/db_xref="taxid: 9606:34812"', and '/db_xref="UniProtKB: P62288"'. The right sidebar contains various navigation and analysis options.

And this is the sequence feature which is highlighted which shows that it is the iso form B is encoded by transcript variant 2 and so on.

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BLAST [®] → *blastn* output → RID-STNC65V104 | Home | Recent Results | Saved Strategies | Help

Format Request Status

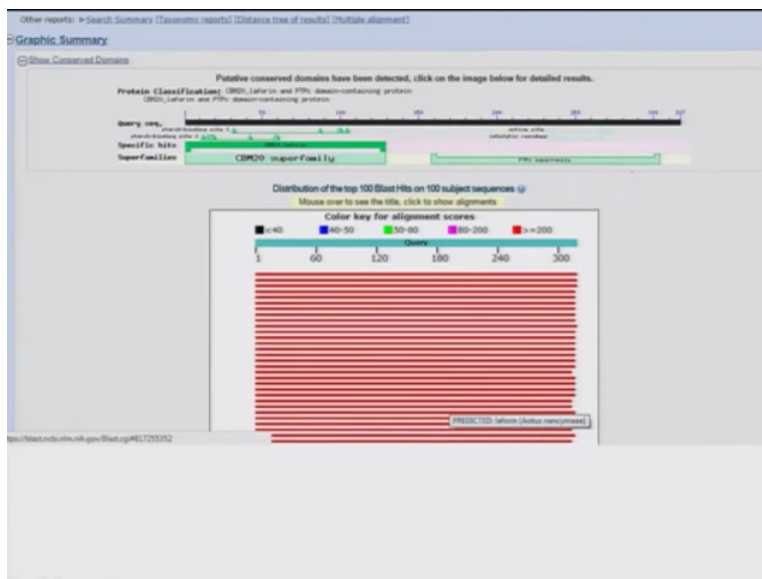
Job Title: refNP_001018051.1 (217 letters)

Request ID	SNC65V104
Status	Searching
Submitted at	Fri Dec 23 02:43:02 2016
Current time	Fri Dec 23 02:43:05 2016
Time since submission	00:00:02

This page will be automatically updated in 2 seconds

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Similarly you can run a blast and what do you find that you find that again this blast would take NP as a query sequence and using the default settings of the search engine when we run blast. This page appears showing that it is running the the job which we have assigned to it and then you get the results again in a similar fashion. How well your query sequence is aligning to the subjects sequence what are the top hits what are organisms it is aligning all those information we can get from here.

(Refer Slide Time: 25:50)

The screenshot shows a web-based multiple sequence alignment tool. The top section is titled "Multiple Alignment Results - ref|NP_001018051.1| (317 letters) - Cobalt RID STNS39HW211 (4 seqs)". Below this, there are tabs for "Descriptions" and "Alignments". The "Descriptions" tab is active, showing a table with columns for "Accession", "Description", and "Links". The table lists four sequences: NP_001018051.1 (Human), NP_001087462.1 (Monkey), NP_034275.2 (Mouse), and NP_001026248.1 (Chicken). Below the table, the "Alignments" tab is visible, showing a compact view of the sequence alignment with conservation scores.

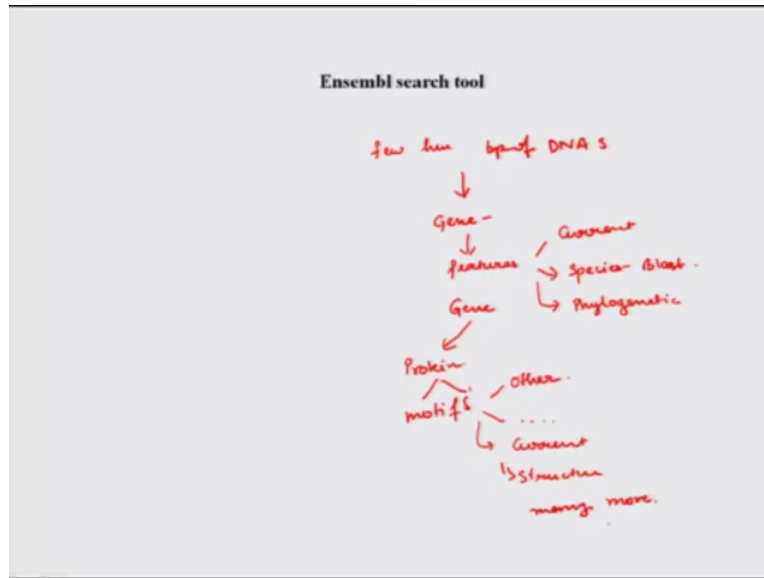
You can also select few sequences for example here I selected the proteins sequence from the humans then monkey then mouse and chicken and then you can see how well it is aligning and you can also identify what all regions are what all regions are more conserved.

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This screenshot shows a detailed view of the multiple sequence alignment. The "Alignments" tab is active, and the "View Format" is set to "Compact". The "Conservation Setting" is set to "2 Bits". The alignment shows four sequences: NP_001018051.1 (Human), NP_001087462.1 (Monkey), NP_034275.2 (Mouse), and NP_001026248.1 (Chicken). The sequences are aligned, and the conservation scores are shown at the end of each line. The alignment shows that the human sequence is not aligning well with the other three sequences, particularly in the last few base pairs.

So you see that in these four groups which I which I have selected mainly most of it is aligning and only like suppose the one of the humans is not aligning to the other three that is monkey, mouse or and the chicken. The few last base pairs are not aligning that well.

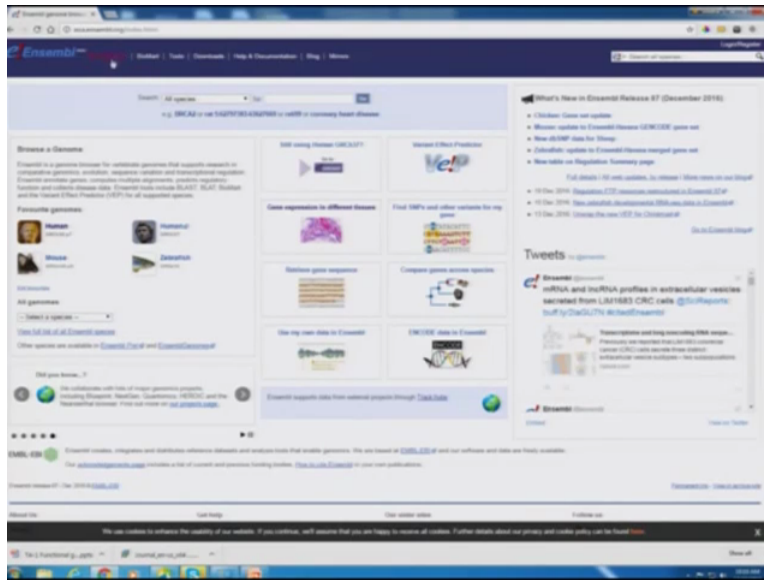
(Refer Slide Time: 26:32)



This is how this huge amount of information we just got from few hundred base pair of the DNA sequence so what all we arrived at that we had few hundred base pairs few hundred base pair of the DNA sequence. From there we arrived to a gene then we looked into what are the features of that gene, we can look into that. What is the current literature about that we looked into in what all species it is present that is by using blast. We can also look into the how conserved that is that is by looking into the phylogenetic tree in and what all organisms it is there. From that gene you can look go to the protein sequence, you can see into what all motives or conserved motives it has how well it is conserved in the other species and so on. What is the current literature about that protein you can look into the structure of that protein and many more.

So this is the power of Bio-informatics and computer science which has been given to the biologists that when you have a small sequence this all you can search it against all the data base present. Now we will look into another search engine that is Ensembl search engine and very quickly we will go we have almost understood what all we can do but let us look into what we can do. So now we will look into another search engine that is Ensembl and we will go online and look into what all information can be fished from that search engine.

(Refer Slide Time: 28:32)



So this is the Ensembl home page and you can do here blast or what are other tools are available similarly you can have your favourite genome selected if you want the gene to be only looked into humans or in zebra-fish or mouse information, similarly you can go for gene expression in different tissues how you gene expresses in different tissues of your organism of your interest, you can compare the gene across the species which we have also done with the help of NCBI using the phylogenetic tree which we created selecting all the sequences.

You can use your own data or you can also upload the data which you have created so depending on your requirement you can search the available search engines for the information and you can explore whatever you wished to. There are various tutorials already available on the YouTube which we can use to do so that is all thank you I hope this session has been somewhat informative for you and which will help you in using these search engines that is all. Thank you.